

6'-*epi*-FORTIMICINS

OBSERVATIONS RELEVANT TO THE MECHANISM OF THE REDUCTIVE AMINATIONS OF KETONES WITH SODIUM CYANOBOROHYDRIDE AND AMMONIUM ACETATE†

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Abstract—Fortimicin B (2) has been converted to 6'-*epi*-fortimicin B (5) and 6'-*epi*-fortimicin A (4) both of which have the same diaminosugar moiety as that present in gentamicin C₂ (3). The syntheses of the 6'-*epi*-fortimicins and their antibacterial activities are reported.

Although reductive amination of 1,2'-di-N-benzyloxycarbonyl-6'-oxo-fortimicin B-4,5-carbamate (17) with sodium cyanoborohydride and ammonium acetate in methanol gave a mixture of 1,2'-di-N-benzyloxycarbonyl-6'-*epi*-fortimicin B-4,5-carbamate (18) and 1,2'-di-N-benzyloxycarbonylfortimicin B-4,5-carbamate (14), attempted reduction of 1,2'-di-N-benzyloxycarbonyl-6'-iminofortimicin B-4,5-carbamate (16), under identical conditions gave only recovered imine. The significance of these results relative to the mechanism of the reductive amination of carbonyl compounds with sodium cyanoborohydride and amine salts is discussed.

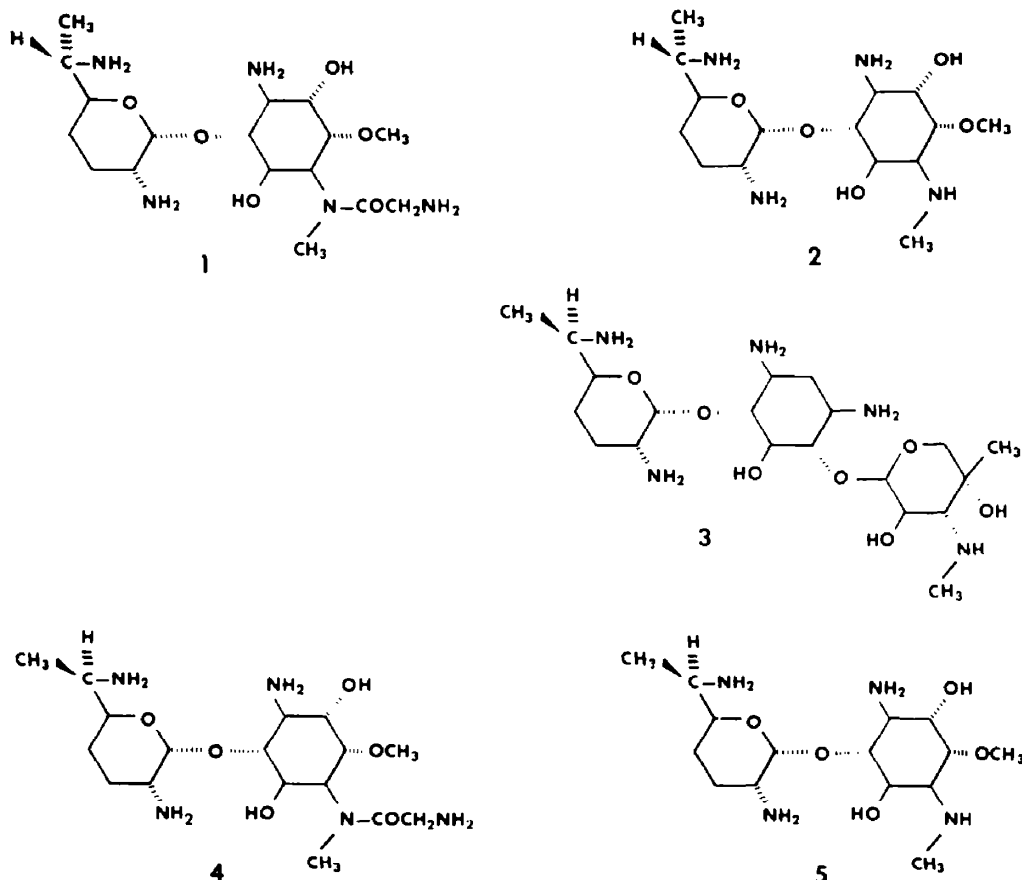
DISCUSSION

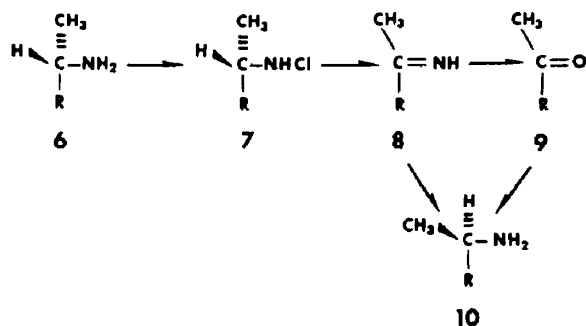
The determination of the structures of fortimicin A (1) and fortimicin B (2)¹ established that the diaminosugar moiety, present in both, differed from that present in gentamicin C₂² (3) only in the configuration of the

secondary carbon (C₆) of the aminoethyl side chain. In the context of a program of chemical modification of the fortimicin antibiotics, carried out with the object of preparing fortimicin derivatives with improved therapeutic properties, we have carried out the syntheses, from fortimicin B, of the 6'-*epi*-fortimicins A and B (4 and 5).

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Our general plan for the preparation of the 6'-*epi*-fortimicins involved preparation of a 6'-oxofortimicin 9





Scheme 1.

by a Ruschig degradation³ (Scheme 1). It was hoped that suitable conditions could be found to effect reductive amination of 9 with the desired stereochemistry to form the 6'-*epi* derivative 10. Our first objective involved conversion of fortimicin B (2) to a derivative with a free 6'-amino group, and with the amino groups at C₁, C₄ and C₂ protected by acyl groups. This was accomplished by first converting fortimicin B to 1,2'-di-N-benzyloxycarbonylfortimicin B (11).⁴ Treatment of the latter with excess N-(2,2,2-trichloroethoxy)phthalimide gave the 4,6'-di-N-(2,2,2-trichloroethoxy) derivative 12 which was detected by thin layer chromatography. The latter was cyclized without purification to 1,2'-di-N-benzyloxycarbonyl-6'-N-(2,2,2-trichloroethoxy)fortimicin B-4,5-carbamate (13) on treatment with sodium bicarbonate in refluxing, aqueous methanol. Cleavage of the 2,2,2-trichloroethoxy group of 13 with zinc in glacial acetic acid gave 1,2'-di-N-benzyloxycarbonylfortimicin B-4,5-carbamate (14). The latter 14 was converted to the 6'-N-chloroamine 15 with N-chlorosuccinimide in methylene chloride. Dehydrohalogenation of 15 with triethylenediamine in dry ethanol gave the 6'-imine 16. Mild acid-catalyzed hydrolysis of the latter 16 gave the 6'-oxo derivative 17. Although the

intermediates, with the exception of the 4,6'-di-N-(2,2,2-trichloroethoxy) derivative and the labile 6'-N-chloro derivative, were purified by chromatography for analysis, the imine 16 and the ketone 17 of adequate purity for further reactions, were prepared from 1,2'-di-N-benzyloxycarbonylfortimicin B (11) without purification at any step.

The imine 16 and the 6'-oxo derivative 17 were characterized by signals in their CMR spectra at 180.7 and 207.9 ppm, respectively, due to their C₆-carbon atoms, and by singlets in their PMR spectra at δ2.05 and δ2.06, respectively, due to their C₆-Me protons.

Reductive amination of the 6'-oxo derivative 17 by the method of Borch *et al.*⁵ gave the 6'-*epi* and 6'-normal amines 18 and 14 in isolated yields of 18% and 12% respectively, based on 1,2'-di-N-benzyloxycarbonylfortimicin B (11). In contrast, attempted reduction of the 6-imine 16 under the same conditions gave recovered imine 16.

To ensure that the failure to effect reduction of the imine 16 was not due to a difference in the pH of the medium from that of the successful reductive amination of the ketone 17⁶ which was carried out under the same conditions, the following experiments were carried out. The pure ketone 17, the pure imine 16, and a 1:1 mixture

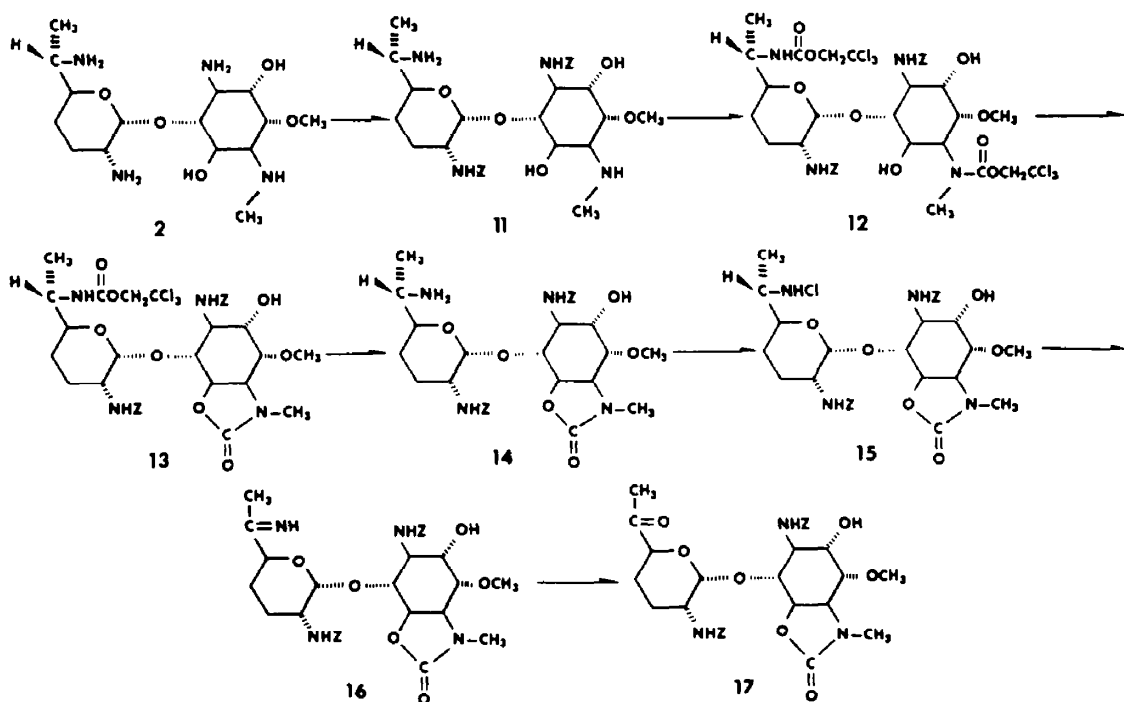
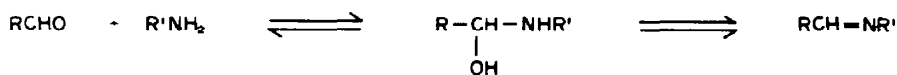


Table 1. Treatment of 1,2'-di-N-benzoyloxycarbonyl-6'-oxo-fortimicin B-4,5-carbamate (17) and 1,2'-di-N-benzoyloxycarbonyl-6'-imino-fortimicin B-4,5-carbamate (16) and a mixture of 16 and 17 with sodium cyanoborohydride and ammonium acetate in methanol

Run	Ketone μg	Imine μg	NaBH_3CN (g)	NH_4OAc (g)	CH_3OH (ml)	Initial ² pH	Final ² pH	% Recovery of Imine μg	Total % Yield Amines μg & μg
Run 1 ¹	0.400	-----	0.060	0.605	7.7	8.0	7.9 (17 hr)	-----	466
Run 2	-----	0.400	0.060	0.605	7.7	7.7	7.9 (17 hr)	933	-----
Run 3 ¹	0.400	0.400	0.120	0.121	15.5	7.9	8.0 (19 hr)	954	707

1. TLC assay showed that reductive amination of the ketone μg was complete within one hour.
2. Apparent pH's determined with a Miller-Markson, Model 80, Digital Mini pH Meter.
3. Total product isolated by extraction, PMR, IR and TLC identical with those of starting material.
4. Isolated by column chromatography as described in the experimental section. Yield based on starting imine μg .
5. The discrepancy in the yields is believed to be due to loss on the chromatography column. The higher yield was obtained by elution with a more polar solvent system. See footnotes 6 and 7.
6. Eluted with a solvent system composed of methylene chloride-ethanol-methanol-concentrated ammonium hydroxide (18:11:0.1). Yield based on starting ketone μg .
7. Eluted with methylene chloride-methanol-concentrated ammonium hydroxide (9:4:0.4). Yield based on starting ketone μg .



Scheme 2.

of ketone and imine were treated in parallel with sodium cyanoborohydride and ammonium acetate in methanol with identical ratios and concentrations of total fortimicin derivatives and reagents (Table 1). The initial apparent pH's of the solutions were between 7.7 and 8.0. The final apparent pH's after 17–19 hr were between 7.9 and 8.0. Tlc determinations established that reductive amination of the ketone 17 was complete within 1 hr for both the reaction of the pure ketone and the reaction carried out with the 1:1 mixture of ketone and imine. Products were isolated after 17–19 hr. As reported above the imine was recovered in 93% yield from the reaction with the pure imine. In addition, the imine 16 was recovered in 95% yield based on starting imine by chromatography of the product mixture obtained from the reaction of the 1:1 mixture of ketone and imine.

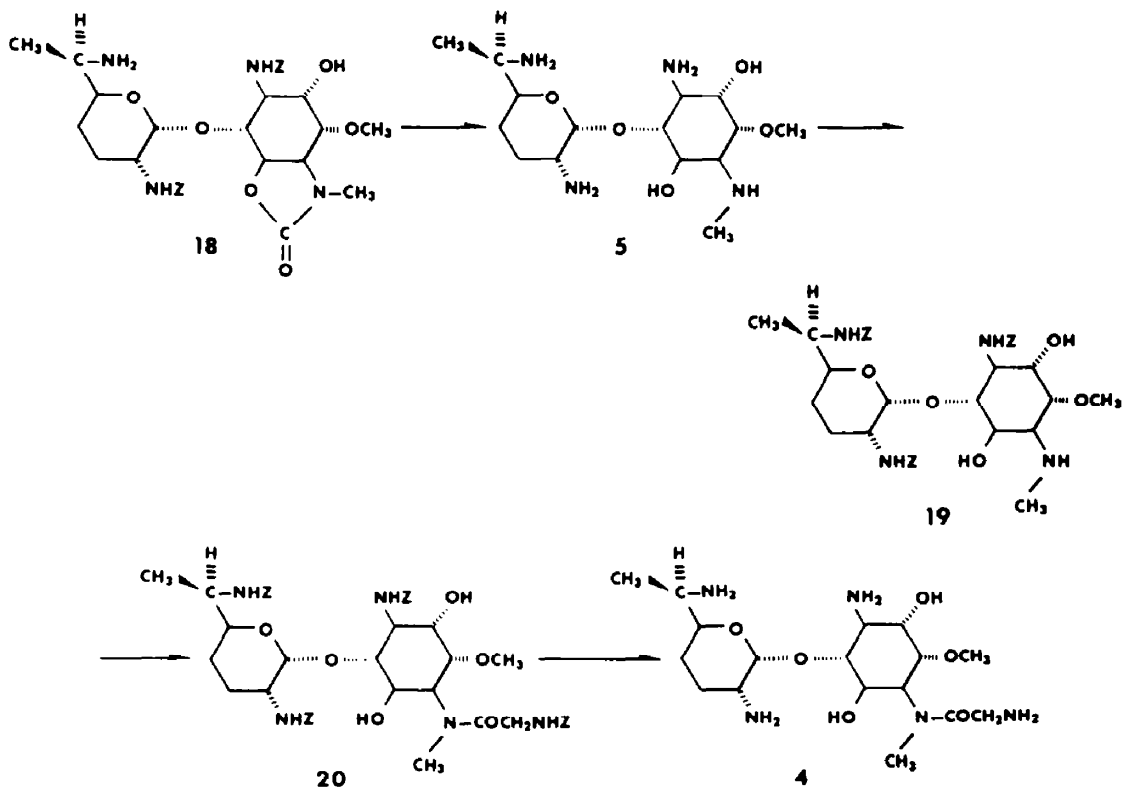
The recovery of the imine from conditions which effect reductive amination of the ketone proves that, contrary to the proposed mechanism⁵ of reductive amination of ketones with sodium cyanoborohydride and ammonium acetate in methanol, the imine 16 is not an intermediate in the reductive amination of the ketone 17. The proposal⁵ that imines were the intermediates reduced in reductive aminations of aldehydes and ketones with sodium cyanoborohydride and amines was based on the fact that the reactions of such carbonyl compounds with amines give rise to imines. It is known, however, that the imines are formed via geminal hydroxyamines⁷ (Scheme 2). Since in the present case the results exclude the imine 16 as the intermediate in the

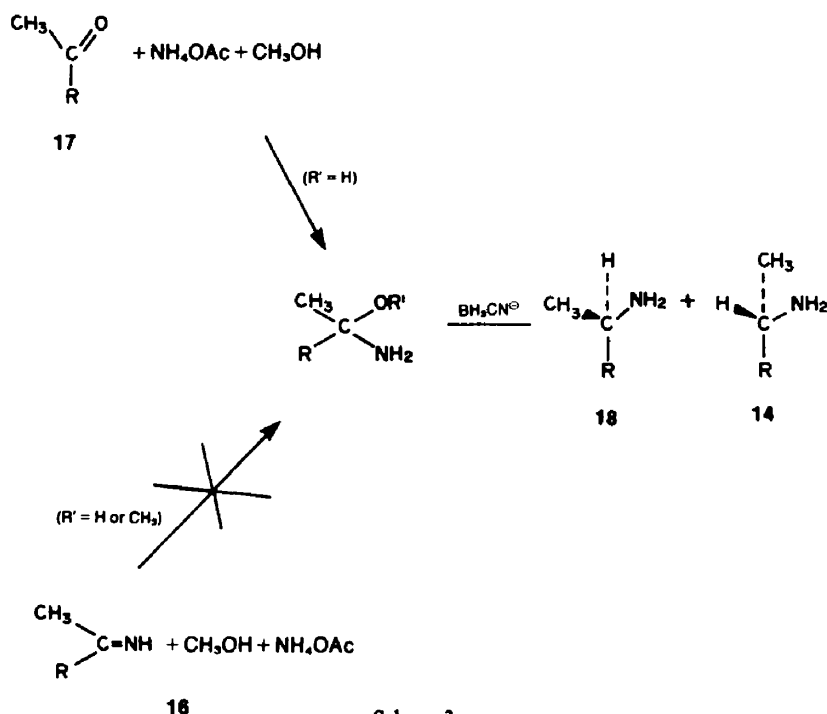
reductive amination of the ketone 17, we propose that the intermediate involved is the geminal hydroxyamine (Scheme 3). Failure of the imine 16 to be reduced under the reductive amination conditions may then be interpreted as a consequence of its failure to undergo conversion to the geminal hydroxyamine or the corresponding geminal methoxyamine (Scheme 3). We thus propose that reductive aminations of aldehydes and ketones with amines and sodium cyanoborohydride may occur, in general, via geminal hydroxyamines rather than imines.

Consistent with this mechanism is the report by Kapanang *et al.*⁸ that bis (methoxymethyl) amines which were prepared and isolated, were quantitatively reduced by sodium cyanoborohydride in methanol to the corresponding dimethylamines. For precedence these authors pointed out the ease of cleavage by nucleophiles of the C–O bonds of carbinolamine ethers.

More favorable stereoselectivity in formation of the desired 6'-*epi* derivative 18 was achieved by direct reduction of the 6'-imine 16 with sodium borohydride which gave the 6'-*epi* amine 18 in 41% yield and the 6'-normal amine 14 in 3% yield based on 1,2'-di-N-benzoyloxycarbonylfortimicin B (11).

Hydrolysis of 1,2'-di-N-benzoyloxycarbonyl-6'-*epi*-fortimicin B-4,5-carbamate (18) with potassium hydroxide gave 6'-*epi*-fortimicin B (5). The latter 5 was converted to 6'-*epi*-fortimicin A (4) by the procedure previously employed for conversion of fortimicin B (2) to fortimicin A (1).⁹ 6'-*epi*-Fortimicin B (5) was converted to 1,2',6'-tri-N-benzoyloxycarbonyl-6'-*epi*-





Scheme 3.

fortimicin B (19) with *N*-(benzyloxycarbonyloxy)succinimide. Acylation of 19 with *N*-(*N*-benzyloxycarbonyl)glycyloxy)succinimide gave 1,2',6',2'-tetra-*N*-benzyloxycarbonyl-6'-*epi*-fortimicin A (20). Catalytic hydrogenolysis of the benzyloxycarbonyl groups of 20 with Pd-C in 0.2 N hydrochloric acid gave 6'-*epi*-fortimicin A (4) as the tetrahydrochloride salt. The latter was converted to the disulfate salt with AG1-X2 (SO₄²⁻-form) anion exchange resin.

The CMR parameters of 6'-*epi*-fortimicin B (5), fortimicin B (2)¹ and gentamicin C₂ (3)¹⁰ are listed in Table 2. CMR spectra were recorded at various pH values in order to determine the pertinent chemical shift changes due to protonation of the amino groups. The data were plotted¹¹ to correlate chemical shifts at highest and lowest pH values. Comparison of the spectra of 5, 2 and

3 was made at the lowest pH to minimize variations in chemical shifts which might be caused by incomplete protonation at higher pH. Comparison of the spectrum of 6'-*epi*-fortimicin B with fortimicin B showed good agreement of the chemical shifts of the aminocyclitol carbons. The chemical shifts of the C₂, C₃, C₄, C₅, C₆ and C₇ carbons of the diaminosugar of 6'-*epi*-fortimicin B were all in excellent agreement with those of gentamicin C₂. In contrast the chemical shifts of the C₂, C₄ and C₇ carbons of 6'-*epi*-fortimicin B differed significantly from those of fortimicin B. That the chemical shift of the anomeric carbon of 6'-*epi*-fortimicin B was in slightly better agreement with that of fortimicin B than with that of the anomeric carbon of the diaminosugar of gentamicin C₂ may reflect the fact that the fortimicins have a common aminocyclitol moiety. This is

Table 2. CMR parameters¹

C	Fortimicin B (2)			6'- <i>epi</i> -Fortimicin B (5)			Gentamicin C ₂ (3)			Free base ³
	pD 1.8	pD 10.3	δ-Shift	pD 1.2	pD 10.2	δ-Shift	pD 2.9	pD 10.1	δ-Shift ²	
1	53.5	53.8		53.5	53.9					
2	65.5	71.2	5.7	65.4	71.2	5.8				
3	74.1	79.9	5.8	74.7	79.9	5.2				
4	58.1	62.8		57.9	62.8					
5	65.6	71.2	4.6	66.5	71.2	4.7				
6	74.2	84.1	9.9	74.1	84.1	9.0				
1'	96.0	102.5	6.5	96.7	102.7	6.0	95.5	101.1	5.6	102.6
2'	51.9	50.6		50.4	50.5		50.4	50.7		51.0
3'	21.5	27.0	5.5	21.8	27.0	5.2	21.9	27.0	5.1	27.1
4'	26.3	27.3		23.9	25.9		23.9	26.0		26.2
5'	71.0	75.1	4.1	69.7	74.4	4.7	69.6	74.4	4.8	74.5
6'	49.4	50.4		49.4	50.0		49.6	50.2		50.3
7'	15.1	18.5	3.4	13.2	18.6	5.4	13.1	19.0	5.9	19.2
NCH ₃	32.3	35.4		32.2	35.4					
OCH ₃	56.1	59.2		57.5	59.2					

¹ 25.2 MHz in D₂O. pD values are uncorrected pH measurements on D₂O solutions.

² Determined from values obtained in these laboratories.

³ Reference 8.

Table 3. *In vitro* antimicrobial activities*

Organism	6'- <i>epi</i> -Fortimicin A (4)	Fortimicin A (1)
<i>Staph. aureus</i> Smith	1.56	0.78
<i>Strep. faecalis</i> 10541	100	50
<i>Enterobacter aerogenes</i>	12.5	3.1
<i>E. coli</i> Juhl	12.5	6.2
<i>E. coli</i> BL-3676	50	25
<i>E. coli</i> 76-2	6.2	3.1
<i>Kleb. pneumoniae</i> 10031	12.5	1.56
<i>Kleb. pneumoniae</i> KY-4262	50	3.1
<i>Providencia</i> 1577	3.1	1.56
<i>Pseudo. aeruginosa</i> BMH #10	3.1	0.78
<i>Pseudo. aeruginosa</i> KY 8512	25	12.5
<i>Pseudo. aeruginosa</i> KY 8516	25	6.2
<i>Pseudo. aeruginosa</i> 209	>100	>100
<i>Pseudo. aeruginosa</i> 27853	50	25
<i>Sal. typhimurium</i> Ed. #9	6.2	3.1
<i>Serratia marcescens</i> #003	6.2	1.56
<i>Shigella sonnei</i> 9290	12.5	6.2
<i>Proteus vulgaris</i> JJ	6.2	3.1
<i>Proteus mirabilis</i> Fin #9	6.2	6.2

* The *in vitro* antibacterial activities were determined by the serial, two-fold dilution method using Mueller-Hinton agar. The antibiotics were assayed as their disulfate salts. Activities are expressed as micrograms of free base per milliliter.

consistent with the observation^{11b} that the chemical shifts of the anomeric carbons of glycosides are sensitive to the nature of the alkyl group bonded to the bridging glycosidic oxygen. It has been suggested^{11b} that the chemical shifts of anomeric carbons are affected by rotamer populations about the C-O-C glycosidic bonds.

The *in vitro* antimicrobial activities of 6'-*epi*-fortimicin A (4) and fortimicin A (1) against sixteen organisms are listed in Table 3. The overall activity of 6'-*epi*-fortimicin A was less than one half that of fortimicin A.

EXPERIMENTAL

General methods. The general methods are described in a related paper.⁴

N-(2,2,2-Trichloroethoxycarbonyloxy)phthalimide

A stirring, ice bath-cooled soln prepared from 65.2 g of N-hydroxyphthalimide, 60.8 g Et₃N and 400 ml N,N-dimethylacetamide was treated dropwise with 93.2 g 2,2,2-trichloroethoxycarbonyl chloride. After addition was complete, stirring was continued in the cold for 1 hr and then at room temp for 16 hr. The mixture was poured into 2.5 l ice water and allowed to stand for 1 hr. The crystalline mass which separated was collected on a filter and washed with water. The solid was dissolved in CHCl₃ and washed with 5% NaHCO₃ aq and dried (MgSO₄). Evaporation of the CHCl₃ gave an oil which crystallized on trituration with hexane to give 105.4 g N-(2,2,2-trichloroethoxycarbonyloxy)phthalimide: m.p. 102-103.5°; IR (CDCl₃) 1815, 1789 and 1745 cm⁻¹; PMR (CDCl₃) δ 4.93 (s, CH₂), 7.87 (m, aromatic). (Found: C, 38.92; H, 1.80; N, 4.08; Cl, 31.99. Calc. for C₁₁H₆NO₃Cl₃: C, 39.02; H, 1.79; N, 4.14; Cl, 31.42%).

1,2'-Di-N-benzyloxycarbonyl-6'-N-(2,2,2-trichloroethoxycarbonyl)-fortimicin B-4,5-carbamate (13)

A soln prepared from 6.2 g of 11,⁴ 10.2 g N-(2,2,2-trichloroethoxycarbonyloxy)phthalimide and 120 ml CHCl₃ was stirred at room temp for 18 hr. The soln was shaken with a mixture of 5% NaHCO₃ aq and CHCl₃. The CHCl₃ layer was separated and evaporated to dryness to give 12 as a bright yellow oil. The oil was heated under reflux for 1.5 hr in a soln prepared from 100 ml MeOH, 20 ml water and 8.5 g NaHCO₃. TLC examination of the soln showed the conversion of 12 to a new, slower moving product which was isolated by CHCl₃ extraction after dilution of the mixture with 5% NaHCO₃ aq. Evaporation of the CHCl₃ gave 10.3 g crude 13 as a glassy residue.

For an analytical sample, 12.0 g product prepared as described above was chromatographed on a column of 400 g silica gel prepared and eluted with a solvent system composed of 1,2-dichloroethane-methanol-conc. ammonium hydroxide (17.2:2.8:0.1) to yield 8.4 g pure 13: [α]_D²⁵ +0.2° (c 1.0, MeOH); IR (CDCl₃) 3457, 3435, 3332, 1760 and 1717 cm⁻¹; PMR (CDCl₃) δ 0.99 (d, C₆-CH₃, J_{6,7} = 6.6 Hz), 2.84 (s, NCH₃), 3.46 (s, OCH₃). (Found: C, 51.40; H, 5.49; N, 6.82; Cl, 12.41. Calc. for C₃₃H₄₃N₄O₁₂Cl₃: C, 51.38; H, 5.30; N, 6.85; Cl, 13.00%).

1,2'-Di-N-benzyloxycarbonylfortimicin B-4,5-carbamate (14)

A suspension prepared from 10.3 g crude 13, 150 ml glacial AcOH and 22 g Zn dust was stirred for 18 hr at room temp. The Zn was removed by filtration. The filtrate was diluted with water and extracted with CHCl₃. The CHCl₃ extract was washed with 5% NaHCO₃ aq and dried (MgSO₄). Evaporation of the CHCl₃ gave 7.20 g residue. A 12.0 g sample thus prepared was chromatographed on a column of 400 g silica gel prepared and eluted with a solvent system composed of 1,2-dichloroethane-methanol-conc. ammonium hydroxide (17.2:2.8:0.1) to give 8.35 g of 14: [α]_D²⁵ +33.2° (c 1.0, MeOH); IR (CDCl₃) 3440, 1750 and 1704 cm⁻¹; PMR (CDCl₃) δ 0.86 (d, C₆-CH₃, J_{6,7} = 6.2 Hz), 2.83 (s, NCH₃), 3.43 (s, OCH₃). (Found: C, 59.41; H, 6.74; N, 8.96. Calc. for C₃₂H₄₂N₄O₁₀: C, 59.80; H, 6.59; N, 8.72%).

1,2'-Di-N-benzyloxycarbonyl-6'-N-chlorofortimicin B-4,5-carbamate (15)

A soln prepared from 2.56 g of 14, 1.56 g N-chlorosuccinimide and 100 ml CH₂Cl₂ was stirred at room temp for 1 hr. Evaporation of the CH₂Cl₂ gave crude 15: IR (CDCl₃) 3553, 3439, 1754 and 1713 cm⁻¹; PMR (CDCl₃) δ 0.902 (d, C₆-CH₃, J_{6,7} = 3.1 Hz), 2.86 (s, NCH₃), 3.46 (s, OCH₃).

Singlets at 2.63 and 2.81 were attributed to the presence of succinimide and N-chlorosuccinimide which could be completely removed by passage of a soln of 15 in MeOH through a column of AG2-X8 (OH) resin packed in MeOH giving the purified 15 the PMR spectrum of which was consistent with its structure.

1,2'-Di-N-benzyloxycarbonyl-6'-iminofortimicin B-4,5-carbamate (16)

A soln of the crude 15, prepared as described,¹² in 200 ml 1% soln of triethylenediamine in anhyd EtOH, previously dried over 3A molecular sieves, was allowed to stand at room temp for 1.3 hr. The soln was shaken with a mixture of CHCl₃ and 5% NaHCO₃ aq. The CHCl₃ layer was separated and washed with 5% NaCl aq. The aqueous layer was washed with CHCl₃. The combined CHCl₃ solns were dried (MgSO₄) and evaporated to

dryness to give 3.0 g white solid. The solid was chromatographed on a column containing 50 ml anion exchange resin, AG2-X8 (OH⁻ form), prepared and eluted with MeOH to give 2.54 g crude 16, free of *N*-chlorosuccinimide and succinimide. For an analytical sample, the crude 16 was chromatographed on a column of 250 g silica gel prepared and eluted with 1,2-dichloroethane-MeOH (18.5:1.5) to give 1.2 g of 16: $[\alpha]_D^{25} + 14.9^\circ$ (c 1.0, MeOH); IR (CDCl₃) 3556, 3434, 1758, 1712 and 1616 cm⁻¹; PMR (CDCl₃) 82.05 (s, C₆-CH₃), 2.86 (s, NCH₃), 3.44 (s, OCH₃). (Found: C, 57.09; H, 5.89; N, 8.01; Cl, 5.54. Calc. for C₃₂H₄₀N₄O₁₀·0.35 CHCl₃: C, 56.92; H, 5.95; N, 8.21; Cl, 5.45%).

Later fractions contained mixtures of 16 contaminated with small amounts of 17.

1,2'-*Di*-*N*-benzyloxycarbonyl-6'-*oxo*fortimicin B-4,5-carbamate (17)

A soln prepared from 0.91 g crude 16, 20 ml 0.4 N HCl and 60 ml THF was allowed to stand at room temp for 18 hr. The soln was shaken with a mixture of CHCl₃ and 5% NaHCO₃ aq. The CHCl₃ portion was separated and washed with water. The aqueous layer was washed with CHCl₃ and the combined CHCl₃ solns were dried (MgSO₄). Evaporation of the CHCl₃ gave 0.794 g solid which was chromatographed on a column (1.6 × 71 cm) of silica gel prepared and eluted with a solvent system consisting of 1,2-dichloroethane-MeOH (18.5:1.5). Fractions containing only the major product were taken to dryness to give 0.396 g of 17: $[\alpha]_D^{25} + 8.6^\circ$ (c 1.0, MeOH); IR (CDCl₃) 3559, 3439, 1760 and 1713 cm⁻¹; PMR (CDCl₃) 82.06 (s, C₆-CH₃), 2.86 (s, NCH₃), 3.45 (s, OCH₃). (Found: C, 59.81; H, 6.40; N, 6.62. Calc. for C₃₂H₃₉N₃O₁₁: C, 59.89; H, 6.13; N, 6.55%).

1,2-*Di*-*N*-benzyloxycarbonyl-6'-*epi*-fortimicin B-4,5-carbamate (18)

(a) A soln prepared from 0.645 g of 17, 0.085 g sodium cyanoborohydride, 0.945 g ammonium acetate and 12.5 ml MeOH was stirred at room temp for 18 hr. The mixture was shaken with a mixture of CHCl₃ and 5% NaHCO₃ aq. The CHCl₃ portion was separated and washed with water. The aqueous layer was in turn washed with CHCl₃. The combined CHCl₃ solns were dried (MgSO₄) and taken to dryness to give 0.570 g white glass. The glass was chromatographed on a column of 40 g silica gel prepared and eluted with a solvent system consisting of CH₂Cl₂-EtOH-MeOH-conc. NH₄OH (18:1:1:0.1). The first fractions eluted gave 0.122 g of 14, identical in all respects with a sample of authentic material.

Continued elution gave fractions containing 0.180 g of 18: $[\alpha]_D^{25} + 36.7^\circ$ (c 1.0, MeOH); IR (CDCl₃) 3437, 1750 and 1702 cm⁻¹; PMR (CDCl₃) 81.02 (d, C₆-CH₃, J_{6,7} = 5.6 Hz), 2.82 (s, NCH₃), 3.44 (s, OCH₃). (Found: C, 58.36; H, 6.68; N, 8.55. Calc. for C₃₂H₄₂N₄O₁₀·H₂O: C, 58.17; H, 6.71; N, 8.48%).

(b) A stirring, ice bath-cooled soln prepared from 5.41 g crude 16, 120 ml MeOH and 24 ml water was treated portionwise with 5.8 g NaBH₄. After addition was complete, stirring was continued in the cold for 2 hr. The mixture was shaken with a mixture of CHCl₃ and 5% NaHCO₃ aq. The CHCl₃ layer was separated and washed with sat NaCl aq. The aqueous solns were washed with CHCl₃ and the combined CHCl₃ solns were dried (MgSO₄) and evaporated to give 4.91 g solid. The solid was chromatographed on a column of 400 g silica gel prepared and eluted with a solvent system composed of CH₂Cl₂-EtOH-MeOH-conc. NH₄OH (18:1:1:0.1). Initial fractions gave 0.276 g of 14 identical in all respects with an authentic sample.

Further elution gave fractions containing 2.73 g of 18 identical with the material obtained by procedure (a) above.

Reductive amination of 1,2'-*di*-*N*-benzyloxycarbonyl-6'-*oxo*-fortimicin B-4,5-carbamate (17) in the presence of 1,2'-*di*-*N*-benzyloxycarbonyl-6'-*imino*-fortimicin B-4,5-carbamate (16)

Recovery of the imine 16. To a stirred soln of 0.400 g of 17, 0.400 g of 16, and 1.21 g ammonium acetate in 15.5 ml MeOH was added 0.120 g sodium cyanoborohydride. Stirring was continued at ambient temp for 19 hr. The product (0.756 g) was isolated by CHCl₃ extraction and chromatographed on a column of 40 g silica

gel packed with a solvent system composed of CH₂Cl₂-MeOH (9:1). Elution with CH₂Cl₂-MeOH (9:1) gave 0.380 g pure 16. Elution with CH₂Cl₂-MeOH-conc. NH₄OH (9:4:0.4) gave 0.281 g of a mixture of 14 and 18.

Attempted reduction of 1,2'-*di*-*N*-benzyloxycarbonyl-6'-*imino*fortimicin B-4,5-carbamate (16) with sodium cyanoborohydride in methanol in the presence of ammonium acetate

Recovery of the imine (16). To a stirred soln of 0.4525 g of 16, 0.6857 g ammonium acetate and 8.75 ml MeOH was added 0.0677 g sodium cyanoborohydride. Stirring was continued for 18 hr. CHCl₃ extraction gave 0.4230 g of recovered 16.

6'-*epi*-Fortimicin B (5)

A soln prepared from 17.8 g of 18 and 125 ml 6 N KOH and 250 ml EtOH, previously purged with N₂, was heated at 80° overnight under N₂. The mixture was cooled in an ice bath and adjusted to pH 7.0 with 2 N HCl. Evaporation of the solvent left a residue which was repeatedly co-distilled with anhyd EtOH to remove residual water. The residue was repeatedly triturated with MeOH. The MeOH washings were filtered and the filtrate was evaporated to dryness to give 11.9 g solid which was chromatographed on a column of 550 g silica gel. Elution with the lower phase of a solvent system prepared from CH₂Cl₂-MeOH-water-conc. NH₄OH (2:2:1:1) gave 8.1 g pure 5: $[\alpha]_D^{25} + 28.4^\circ$ (c 1.0, MeOH); PMR (D₂O, pD 11.02) 81.54 (d, C₆-CH₃, J_{6,7} = 7.5 Hz), 2.86 (s, NCH₃), 3.94 (s, OCH₃), 5.55 (d, H₁, J_{1,2} = 3.6 Hz); mass spectrum, *m/e* 348.2391 (M⁺). Calc. for C₁₅H₁₂N₄O₅: 348.2373; sugar fragment *m/e* 143.1190. Calc. for C₇H₁₃N₂O 143.1184; aminocyclitol fragment *m/e* 207.1350. Calc. for C₈H₁₉N₂O₄ 207.1345.

1,2,6'-*Tri*-*N*-benzyloxycarbonyl-6'-*epi*-fortimicin B (19)

A stirring, ice bath-cooled soln prepared from 8.88 g of 5, 260 ml MeOH and 130 ml water was treated with 21.0 g *N*-(benzyloxycarbonyloxy)succinimide. Stirring was continued in the cold for 3 hr and then at room temp for 18 hr. The mixture was added to 11 5% NaHCO₃ aq and extracted with CHCl₃. The CHCl₃ extract was dried (MgSO₄) and evaporated to dryness to give 20.7 g residue. The residue was chromatographed on a column of 750 g silica gel prepared and eluted with a mixture of 1,2-dichloroethane-EtOH-conc. NH₄OH (18:2:0.1) to give 13.8 g of 19: $[\alpha]_D^{25} + 24.7^\circ$ (c 1.0, MeOH); IR (CDCl₃) 3554, 3434, 3334 and 1708 cm⁻¹; PMR (CDCl₃) 81.02 (d, C₆-CH₃, J_{6,7} = 6.4 Hz), 2.34 (s, NCH₃), 3.42 (s, OCH₃). (Found: C, 62.13; H, 6.77; N, 7.37. Calc. for C₃₉H₅₀N₄O₁₁: C, 62.39; H, 6.71; N, 7.46%).

1,2,6,2'-*Tetra*-*N*-benzyloxycarbonyl-6'-*epi*-fortimicin B (20)

A soln prepared from 12.8 g of 19, 5.75 g *N*-(benzyloxycarbonyl)glycyloxy)succinimide and 450 ml THF was stirred for 24 hr at room temp. An additional 0.85 g *N*-(benzyloxycarbonyl)glycyloxy)succinimide was added and stirring was continued for 18 hr. The THF was evaporated and the resulting residue was taken up in CHCl₃. The CHCl₃ soln was washed with 5% NaHCO₃ aq and dried (MgSO₄). Evaporation of the CHCl₃ gave 17.1 g solid which was chromatographed on a column of 750 g silica gel prepared and eluted with EtOAc. Fractions containing the major component were evaporated to give 11.8 g of 20: $[\alpha]_D^{25} + 74.1^\circ$ (c 1.0, MeOH); IR (CDCl₃) 3552, 3415, 1713 and 1637 cm⁻¹; PMR (CDCl₃) (rotamers) 81.78 (d, C₆-CH₃, J_{6,7} = 3.9 Hz), 2.82, 2.98 (NCH₃, major, minor respectively), 3.26 (s, OCH₃). (Found: C, 62.32; H, 6.30; N, 7.34. Calc. for C₄₀H₅₀N₅O₁₄: C, 62.48; H, 6.31; N, 7.43%).

6'-*epi*-Fortimicin A (4)

A soln prepared from 2.83 g of 20 in 240 ml 0.2 N HCl in MeOH was hydrogenated under 3 atmospheres H₂ for 4 hr in the presence of 2.8 g 5% Pd-C. The catalyst was removed by filtration and the filtrate was evaporated to leave a solid. Excess HCl was removed by repeated codistillation with MeOH to give 1.70 g of 4 isolated as the tetrahydrochloride salt: $[\alpha]_D^{25} + 77.7^\circ$ (c 1.0, MeOH); IR (KBr) 1646 cm⁻¹; PMR (D₂O, pD 2.39) 81.78 (d, C₆-CH₃, J_{6,7} = 7.8 Hz), 3.60 (s, NCH₃), 3.96 (s, OCH₃); mass

spectrum, *m/e* 405.2596 (M^+). Calc. for $C_{17}H_{35}N_5O_6$ 405.2587; sugar fragment *m/e* 264.1553. Calc. for $C_{10}H_{22}N_3O_3$ 264.1559; aminocyclitol fragment *m/e* 143.1209. Calc. for $C_7H_{13}N_2O$ 143.1184.

The disulfate salt of 4 was prepared by passing a water soln of 6.61 g 6'-*epi*-fortimicin A tetrahydrochloride through a column (2.5×32 cm) of an anion exchange resin, AG1-X2 ($SO_4 = \text{form}$). Fractions containing the antibiotic were taken to dryness to give 7.0 g of 4 as the disulfate salt: $[\alpha]_D^{25} + 72.4^\circ$ (*c* 1.0, water); IR (KBr) 1646 cm^{-1} ; PMR (D_2O , pD 4.2) δ 1.75 (d, C_6-CH_3 , $J_{6,7} = 6.9 \text{ Hz}$), 3.58 (s, NCH_3), 3.94 (s, OCH_3); mass spectrum, *meas.* 405.2596 (M^+). Calc. for $C_{17}H_{35}N_5O_6$ 405.2587. (Found: C, 31.67; H, 6.65; N, 11.47. Calc. for $C_{17}H_{39}N_5O_{14}S_2 \cdot 2H_2O$: C, 32.01; H, 6.80; N, 10.98%.)

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